

A PARAMETRIC STOCHASTIC MODEL OF BONE GEOMETRY

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The aim of the present study is to develop a parametric bone modelling algorithm which takes into account bone microarchitecture. This approach allows to generate hematopoietic bone segment phantoms based on literature-derived micro- and macro dimensions. We propose a method for subdividing bones into small segments which can be described by simple geometric shapes filled with a stochastically generated rod-like model of the trabecular structure with appropriate voxel resolution. This approach avoids the disadvantages of non-parametric individual modelling based on computer tomography scans. The parametric method allows the simulation of individual variability in bone-specific dimensions. The model presented in this paper will be used to describe the geometry of hematopoietic sites, which in turn will serve as a basis for calculating the doses of irradiation of the hematopoietic cells of the bone marrow from the incorporated beta-emitters.

Keywords: micro- and macro- structure of the trabecular bone; stochastic modelling; voxelization.

Introduction

The bones of the human skeleton are composed of spongiosa covered with a compact bone layer, or cortical bone (Fig. 1). Spongiosa is a porous structure with friable lying bone trabeculae permeating bone marrow – the soft tissue of the internal bone cavity, which includes hematopoietic cells (active bone marrow).

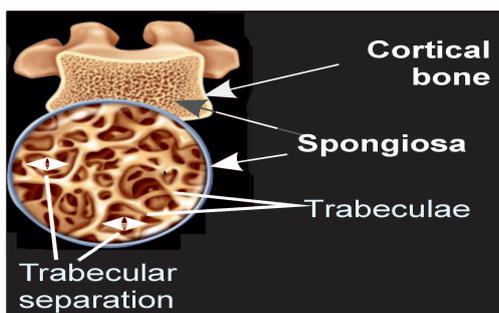


Fig. 1. Outer cortex and internal spongy structure (trabecular bone) of the human lumbar vertebra

Skeleton geometry modelling is widely used in biomedical research, for example, in studying the mechanical properties of bone in traumatology and implantation [1]; in studies of relationships between bone morphology and different diseases (osteoarthritis, osteoporosis) [2]; in radiation dosimetry [3–5], and so on. We are interested in the latter problem.

Different tasks require different levels of geometric details and different degrees of realism. For example, very realistic individual models of a patient's bone shape are created for medical examination with gamma irradiation (computer tomography, cyber knife,

SPECT systems). However, there is no need to build a material microstructure model; both cortical and sponge bones can be considered as homogeneous structures in terms of photon transport [3]. Bone dosimetry for alpha emitters doesn't require a description of the bone shape (due to short pathlength of α -particles). The dimensions of a bone segment model are usually not larger than 2 mm [4]. However, it is extremely important to describe the trabecular structure at the sites of radionuclide localization (for instance, trabecula surfaces and adjacent intertrabecular space) as accurately as possible.

Dose estimation in active marrow exposed to bone-seeking beta-emitters, such as ^{89}Sr or $^{90}\text{Sr}/^{90}\text{Y}$ (energies of electron emission are 0 – 1,5 MeV and 0 – 2,4 MeV, respectively), is an important task in bone dosimetry. Monte Carlo simulations of electron – photon transport to calculate the active marrow doses are based on the geometrical modelling of bone structures [4, 5, 19]. The description of spongy bone microarchitecture in the model is important due to high probability of low-energy electron emission. The mean free paths of the electrons of the $^{90}\text{Sr}/^{90}\text{Y}$ spectra (in the continuous-slowing-down approximation) are in the range of 1–3 mm, depending on the density of the bone media. These dimensions are comparable to the thickness of the cortical layer of skeletal bones. Therefore, the evaluation of the contribution of contaminated cortical bone into bone marrow dose relies on the accurate description of cortical thickness. The maximum free path (for electrons yielding with maximum energy) in a spongy bone can reach 5–9 mm. These dimensions are comparable to the linear dimensions of many bones of the skeleton. Thus, the model geometry should consist of accurate descriptions of the fine structure of spongiosa and cortical bone thickness, as well as descriptions of macro-dimensions of the bone.

New computer tomography (CT)-based methods are widely applied to developed computational phantoms [5], among which there are phantoms designed for bone dosimetry of beta-emitting radionuclides [6 – 8]. Such phantoms were developed using a combination of micro- and macro- CT images of human bones. The advantage of this CT-based method lies in the high realism of the complex bone shape description, as well as the possibility of an adequate description of bone-specific microstructure. However, the method has a number of disadvantages, such as:

- The computations are laborious and expensive.
- A detailed scan of the whole skeleton takes a long time and results in high radiation exposure. Cadavers are commonly used, associated with organizational difficulties.
- A single cadaver based model needs not be representative and does not allow the estimation of uncertainties associated with the variability of human anatomy.
- The voxel dimensions of the model should be at least twice as small as the dimensions of the imitated structural components to allow accurate simulation of electron-photon transport. In practice, the voxel resolution of the model is limited by computational power. For example, the voxel sizes of the models describing large bone segments [6–8] were restricted to 100 – 200 μm , despite the fact that the thickness of the trabeculae (the modeled structural elements) ranges from 40 to 400 μm . Inadequate voxelization can lead to errors in dose calculations. In theory, it is possible to change the voxelization when increased computing power is available, but in practice this is difficult to implement, since it would be necessary to repeat the analysis and alignment of CT and μCT images.
- Cortical bone thickness is fixed based on the CT, however CT resolution (about 1mm) is insufficient to determine cortical thickness for most of bones (for example, about 0,4mm in vertebral bodies [9]).

Moreover, high individual variability of bone shapes and macro-dimensions negates the advantages of the high realism of the model of a single (or a population-average) body.

The aim of the present study is to develop an algorithm of parametric bone modelling, which allows the generation of phantoms of hematopoietic bone segments based on literature-derived micro- and macro dimensions.

We propose an approach that permits easy subdivision of bones into small segments which can be described by simple-shape geometric figures with appropriate voxel resolution. The parameters of the proposed cadaver-free model can be obtained from publications on morphometry and hystomorphometry widely presented in the biomedical literature. The proposed cadaver-free method avoids the disadvantages of non-parametric individual modelling based on CT scans. The parametric approach allows the simulation of individual variability of bone-specific dimensions.

1. Basic Assumptions

We will proceed from the following assumptions.

1. According to preliminary estimates [10], variations in the geometric shape of bones with similar linear dimensions do not introduce a significant error ($<5\%$) in the calculation of doses. Therefore, it is possible to describe the shape of the bone with simple geometric figures (stylized phantoms).

2. The cortical layer is considered as a homogeneous and isotropic substance located between two surfaces, one of which is the outer boundary of the stylized phantom, and the other is separated from it inside the phantom by a distance equal to the cortical thickness. Cortical thickness ($Cb.Th$) is a parameter of the model, which can be estimated from literature data.

3. The trabecular bone can be described by an isotropic stochastically deformed network structure, with beams randomly varying in width. This model can be parametrized using standard literature-derived indicators of bone microarchitecture, such as:

- mean trabecular thickness ($Tb.Th$) and its intra-specimen variability;
- average distance between the trabeculae – trabecular separation ($Tb.Sp$) and its intra-specimen variability; and
- average bone volume – to – total volume ratio of the entire cancellous bone (BV/TV), and the range of its admissible values.

4. The generated analytical model must be voxelized at a given resolution.

2. The Bone Shape Model

The following geometric objects are used as the stylized phantoms designed to describe the *shape* of bones:

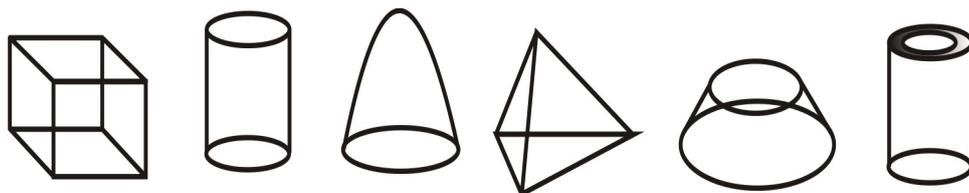


Fig. 2. Stylized phantoms

The parameters of these figures are known from the literature and are specified by the user.

For example, a fragment of a rib body can be described by a rectangular parallelepiped, as shown in Fig. 3. Standardized morphometric indices, such as h and w (see, for example, [11]) are averaged over the entire length of the edge and are attributed to the height and width of the model parallelepiped. The length of the segment of the edge is chosen on the order of 3 lengths of the maximum range of electrons (in the example presented, 3 cm). The average value of the cortical thickness is also taken on the basis of the literature data [12]. Similarly, the neck of the hip is modelled by a cylinder, the head of the hip and humerus is half-ellipsoidal, the wings of the sacrum are triangular, the region of the spines of the femur and humerus is truncated cones, the acetabulum is a double cylinder, etc.

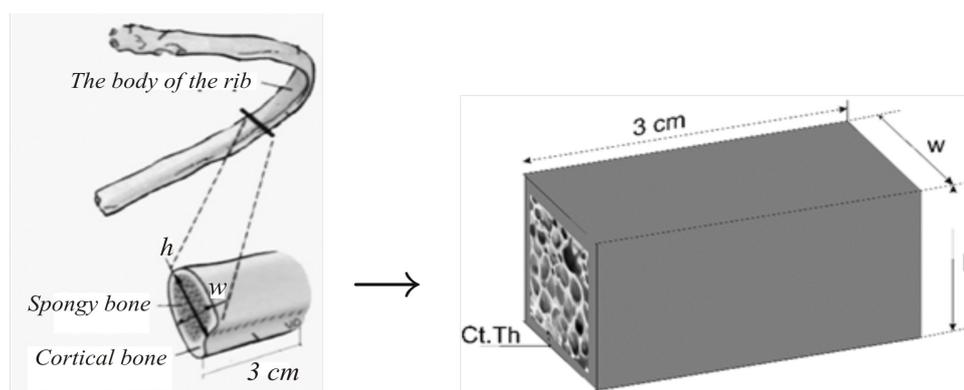


Fig. 3. An example of creating a stylized rib phantom

The main microparameter at this stage is $Cb.Th$. The stylized model of the shape of the bone and its cortical layer encompasses a region of space filled with a trabecular structure. The trabecular structure of the bone is simulated by immersing the phantom in an enclosing rectangular parallelepiped (carcass), and drawing the main parameters, as discussed below.

3. A Stochastic Model of the Trabecular Bone Structure

The microstructure of the trabecular bone is modelled in four stages: construction of the framework, randomization of the framework nodes, stochastic modelling of rod-like bone structures, and calibration of the model.

3.1. Construction of the Frame

The main microparameters at this stage are trabecular separation ($Tb.Sp$) and trabecular thickness ($Tb.Th$). In a simple initial frame with cubic cells $Tb.Sp$ will be equal to the length of the edges of cells, or conditional trabeculae. According to data from the literature [13–15], $Tb.Sp$ can be considered a normally distributed value, while $Tb.Th$ has a lognormal distribution.

We take the correct frame in R^3 composed of rectangular parallelepipeds with edges parallel to the coordinate axes $K = \{(t_1, t_2, t_3) : a_i \leq t_i \leq b_i, i = 1, 2, 3.\}$ to construct the framework (Fig. 4, left). The dimensions of the frame are chosen so that the figures of the stylized phantoms (F) are completely immersed in them: $VK \geq VF$.

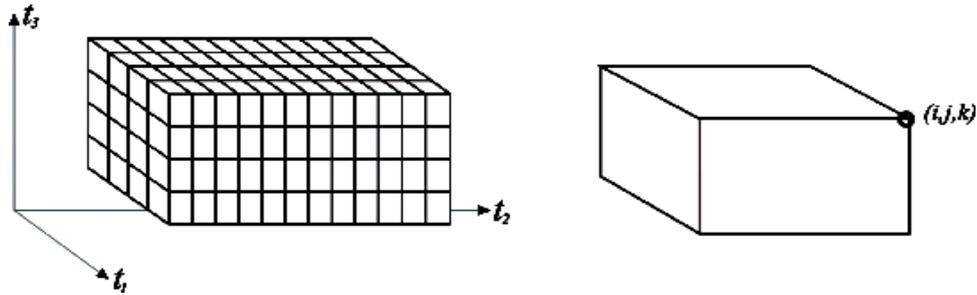


Fig. 4. The framework for microstructure modelling (left) and grid cell notation (right)

Dividing abscissa (t_1), ordinate (t_2) and applicate (t_3) into m, n and l parts, respectively, the frame nodes can be denoted as follows:

$$a_1 = t_1^0 < t_1^1 < \dots < t_1^n = b_1, \quad a_2 = t_2^0 < t_2^1 < \dots < t_2^m = b_2, \quad a_3 = t_3^0 < t_3^1 < \dots < t_3^l = b_3,$$

where each of the frame nodes is denoted by the ordered number triplet (i, j, k) , corresponding to the node coordinate $M_{ijk} = (t_1^i, t_2^j, t_3^k)$. The notation $M_{ijk} = (t_1^i, t_2^j, t_3^k)$ will always refer to a grid cell as shown in Fig. 4 (right), so that grid cell numbers range from $(1, 1, 1)$ to (n, m, l) .

The parameters of the framework partitioning, (i.e., the values $\Delta t_s^i = t_s^{i+1} - t_s^i$, $i = 1, 2, 3$, $s = 0, 1, 2, \dots, N_i$, $N_1 = n, N_2 = m, N_3 = l$,) are determined by the characteristic distances between the trabeculae in the horizontal, vertical and frontal directions.

As a first approximation, we can assume that the values $\Delta t_s^i = t_s^{i+1} - t_s^i$ are sufficiently close to $Tb.Sp$ – the characteristic distance between the trabeculae, so that the horizontal, vertical and frontal dimensions of the frame parallelepiped are multiples of this quantity.

3.2. Node Randomization

Let $\xi = \begin{Bmatrix} \xi_1 \\ \xi_2 \\ \xi_3 \end{Bmatrix}$ denote a Gaussian random variable with the zero vector of averages and the covariance matrix K_ξ

$$K_\xi = \begin{pmatrix} \sigma_1^2 & k_{12} & k_{13} \\ k_{12} & \sigma_2^2 & k_{23} \\ k_{13} & k_{23} & \sigma_3^2 \end{pmatrix}.$$

It should be noted that we assume the node permutations are mutually independent events. Each permuted node \bar{M}_{ijk} can be expressed in terms of its initial state $M = M_{ijk}$ according to the following rule (Fig. 5): $\bar{M} = M + \xi^M \Rightarrow \bar{t}_s^{ijk} = t_s^{ijk} + \xi_s^{ijk}$, $s = 1, 2, 3$

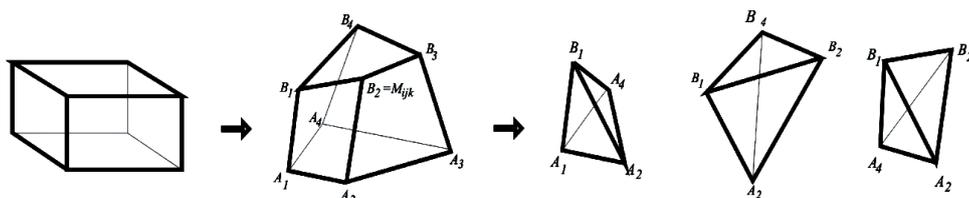


Fig. 5. Transformation of the framework elements and the volume calculation

Thus, the transformed framework is based on the natural transformation of the original edges in accordance with the transformed node positions. For example, an edge initially linking the nodes M_{ijk} and $M_{i+1,jk}$ will turn into an edge linking the mapped nodes (\bar{M}_{ijk} and $\bar{M}_{i+1,jk}$). The randomization parameters (that is, the values σ_i^2 , k_{ij}) are determined by the scattering and correlation relationships of the value $Tb.Sp^1$. As the first approximation, we can assume that $\sigma_i^2 = \sigma^2$, $k_{ij} = 0$.

3.3. Parameters of the Modified Framework

If $A(t_1^A, t_2^A, t_3^A)$ and $B(t_1^B, t_2^B, t_3^B)$ are arbitrary adjacent nodes, then the length of corresponding edge can be expressed as follows:

$$|\bar{A}\bar{B}| = \sqrt{(t_1^B + \xi_1^B - t_1^A - \xi_1^A)^2 + (t_2^B + \xi_2^B - t_2^A - \xi_2^A)^2 + (t_3^B + \xi_3^B - t_3^A - \xi_3^A)^2}.$$

For example, the length of transformed abscissa-directed (frontal) edges ($M_{ijk}M_{i+1jk} \rightarrow \bar{M}_{ijk}\bar{M}_{i+1jk}$) can be calculated as

$$|\bar{M}_{ijk}\bar{M}_{i+1jk}| = \sqrt{(t_1^{i+1} + \xi_1^{i+1,jk} - \xi_1^{ijk} - t_1^i)^2 + (\xi_2^{i+1,jk} - \xi_2^{ijk})^2 + (\xi_3^{i+1,jk} - \xi_3^{ijk})^2}.$$

The length of transformed ordinate-directed (lateral) edges, $M_{ijk}M_{ij+1k} \rightarrow \bar{M}_{ijk}\bar{M}_{ij+1k}$, and applicate-directed (vertical) edges, $M_{ijk}M_{ijk} \rightarrow \bar{M}_{ijk}\bar{M}_{ijk+1}$ edges can be calculated similarly.

Let $\Delta t_s^{p_s}, p_s = 1, 2, \dots, N_s, s = 1, 2, 3$ denote the distance between the parallel edges of the parent framework, where $s = 1$ corresponds to the frontal direction; $s = 2$ to the lateral direction; and $s = 3$ to the vertical direction. The vector of node permutations (for example, in the frontal direction),

$$\Delta \xi = \left\{ \begin{array}{c} \Delta \xi_1 \\ \Delta \xi_2 \\ \Delta \xi_3 \end{array} \right\} = \left\{ \begin{array}{c} \xi_1^{i+1jk} - \xi_1^{ijk} \\ \xi_2^{i+1jk} - \xi_2^{ijk} \\ \xi_3^{i+1jk} - \xi_3^{ijk} \end{array} \right\},$$

is the Gaussian [13] random variable with the zero vector of averages and the covariance matrix

$$K_{\Delta \xi} = \left(\begin{array}{ccc} \sigma_{1(i+1jk)}^2 + \sigma_{1(ijk)}^2 & k_{12}^{\Delta \xi} & k_{13}^{\Delta \xi} \\ k_{12}^{\Delta \xi} & \sigma_{2(i+1jk)}^2 + \sigma_{2(ijk)}^2 & k_{23}^{\Delta \xi} \\ k_{13}^{\Delta \xi} & k_{23}^{\Delta \xi} & \sigma_{3(i+1,jk)}^2 + \sigma_{3(ijk)}^2 \end{array} \right),$$

where $k_{pq}^{\Delta \xi} = k_{pd}^{i+1jk} + k_{pd}^{ijk}$, $p, q = 1, 2, 3$. A similar expression can be easily written for other two directions. If the permutation scattering is significantly smaller than the typical

¹Here it is useful to bear in mind the following circumstance: the average value of the intertrabecular space is very unstable ([11]) and can vary within fairly wide limits. Therefore, the adjustment and selection of the values for the parameter $Tb.Sp$ take into account the behaviour of the ratio of the volume of bone tissue (BV) to the total bone volume (TV). The desired value of this ratio is set by the user as a range of possible values before starting the simulation. If the resulting value of the BV / TV ratio differs from the desired value slightly – the parameters $Tb.Sp$ are accepted. Otherwise, they are adjusted so that the BV / TV ratio falls within the desired range.

size of parent framework structures, $\sigma_{p(ijk)}^2 \ll (\Delta t_p^{ijk})^2$, then a first approximation of the average distance between edges of the transformed framework can be calculated as² :

$$M(\Delta l_{ijk}) = \Delta t_1^i + \left(\frac{\sigma_1^{i+1jk}}{\Delta t_1^i}\right)^2 + \left(\frac{\sigma_1^{ijk}}{\Delta t_1^i}\right)^2 + \left(\frac{\sigma_2^{i+1jk}}{\Delta t_1^i}\right)^2 + \left(\frac{\sigma_2^{ijk}}{\Delta t_1^i}\right)^2 + \left(\frac{\sigma_3^{i+1jk}}{\Delta t_1^i}\right)^2 + \left(\frac{\sigma_3^{ijk}}{\Delta t_1^i}\right)^2.$$

This relationship allows us to specify the parameters of the original framework in order to obtain the desired parameters of the modified one.

3.4. Cell Volume of the Transformed Frame

Let $A_1A_2A_3A_4B_1B_2B_3B_4$ be the nodes of the transformed frame for a given cell (ijk) (Fig. 5)³. The cell can be subdivided into two prisms, $A_1A_2A_4B_1B_2B_4$ and $A_2A_3A_4B_2B_3B_4$, and each of the prisms can be further subdivided into 3 triangular pyramids as shown in Fig.5 to simplify the volume calculation.

Pyramid volume can be calculated as $V = \frac{1}{6}|(\vec{a}, \vec{b}, \vec{c})|$, where $(\vec{a}, \vec{b}, \vec{c})$ is the triple scalar product of the reference vectors of a pyramid.

Assuming (as above) $\sigma_{p(ijk)}^2 \ll (\Delta t_p^{ijk})^2$ and considering only those elements of triple scalar product whose degree of smallness does not exceeded $\sigma_{p(ijk)}^2$, the volume of the pyramid can be calculated as follows⁴:

$$\tilde{V} = \frac{1}{6}((t_1^i - t_1^{i-1})(t_2^j - t_2^{j-1})(t_3^k - t_3^{k-1}) + (\xi_1^{ij-1k-1} - \xi_1^{i-1j-1k-1})(t_2^j - t_2^{j-1})(t_3^k - t_3^{k-1}) + (\xi_2^{ijk-1} - \xi_2^{ij-1k-1})(t_1^i - t_1^{i-1})(t_3^k - t_3^{k-1}) + (\xi_3^{ij-1k} - \xi_3^{ij-1k-1})(t_1^i - t_1^{i-1})(t_2^j - t_2^{j-1})).$$

3.5. Modelling of Stem-Like Bone Structures

We assume that a single trabecula is located on every edge of the modified frame (Fig. 6). The distribution of trabecular thicknesses within most bone samples is somewhat asymmetric and can be approximated by a lognormal distribution [14,15]. In each modified node \bar{M}_{ijk} , we generate a random variable R_{ijk} with given characteristics $M(R_{ijk}) = R$, $\sigma^2(R_{ijk}) = \sigma^2$. If \bar{M}_{i+1jk} is a node adjacent to the node \bar{M}_{ijk} , then the trabecula will be modeled as a truncated cone with the axis $\bar{M}_{ijk}\bar{M}_{i+1jk}$ and spherical surfaces of radii R_{ijk} and R_{i+1jk} respectively at the bases.

The trabecula volume (Fig. 6) can be calculated in 4 steps:

1. Calculation of the conical frustum volume (the radii of the bases are (R_1, R_2) and the length is L):

$$V(L, R_1, R_2) = \frac{1}{3}\pi L(R_1^2 + R_1R_2 + R_2^2), \quad L = \sqrt{l^2 - (R_{i+1jk} - R_{ijk})^2 - (R_1 - R_2)^2},$$

$$R_1 = \frac{R_{i+1jk}\sqrt{l^2 - (R_{i+1jk} - R_{ijk})^2}}{l}, \quad R_2 = \frac{R_{ijk}\sqrt{l^2 - (R_{i+1jk} - R_{ijk})^2}}{l}.$$

²Hereinafter, all equations are presented for frontal-directed perturbations. Lateral- and vertical-directed perturbations can be described by direct analogy.

³The node B_2 in the current notation corresponds to \bar{M}_{ijk}

⁴It should be noted that the average volume of randomly transformed frameworks is practically equal to the volume of the parent framework. In contrast, the average edge lengths become larger after transformation.

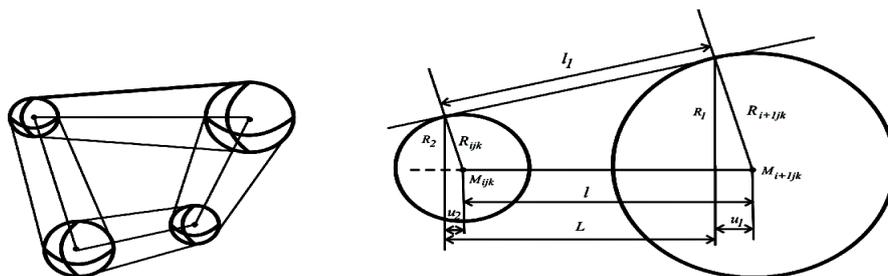


Fig. 6. Trabecula volume calculation

2. Calculation of the volume of the spherical segments: $V_{S_i} = \pi H^2(R - \frac{1}{3}H)$ where H can be computed for two nodes using the parameters shown in the Fig.6. For the left node ($R_{ijk} < R_{i+1jk}$): $H_2 = R_{ijk} + u_2$, $u_2 = \sqrt{R_{ijk}^2 - R_2^2}$, and for the right node ($R_{ijk} < R_{i+1jk}$): $H_1 = R_{i+1jk} - u_1$, $u_1 = \sqrt{R_{i+1jk}^2 - R_1^2}$.

3. Subtraction of the volume of the supplementing spherical segments $V(L, R_1, R_2) - V_{S_i}$.

4. Calculation of the overall trabecular volume as a sum of volumes of conoidal structures (after subtraction of the supplementing spherical segments) and spherical structures at the nodes of the transformed grid.

4. Model Calibration

It should be noted that $Tb.Sp$ is an uncertain parameter and if adopted in the rod-like model it may lead to incorrect estimation of the spongioza density. In other words, this model can only be considered the first approximation. It is therefore important to fit a criterion of adequacy, where an acceptable range for the volume ratio BV/TV is adopted (see above footnote 1), in order to obtain model volume ratios corresponding to values observed in reality. For this purpose, we have developed a multi-step procedure for calibrating $Tb.Sp$ values. The calibration is performed by changing the average distance between nodes at each step until the model ratio $(BV/TV)_i$ is equal to the calibration value $(BV/TV)_{Calibr}$ specified by the user before starting the simulation.

The iterative calibration algorithm is as follows. An additional coefficient ($Tb.Sp.Factor$) is introduced into the formula for calculating the distance between nodes:

$$\Delta t_s^i = r_i^{s+1} - r_i^s = Tb.Sp.Factor \cdot Tb.Sp.$$

The perturbation vector is also multiplied by this coefficient:

$$\Delta \xi_{i+1} = \begin{pmatrix} \Delta \xi_1^{i+1} \\ \Delta \xi_2^{i+1} \\ \Delta \xi_3^{i+1} \end{pmatrix} = Tb.Sp.Factor \cdot \begin{pmatrix} \Delta \xi_1^i \\ \Delta \xi_2^i \\ \Delta \xi_3^i \end{pmatrix}.$$

At the first calibration step, $Tb.Sp.Factor$ is assumed to be equal to 1. We model the cellular structure, obtain the model ratio $(BV/TV)_i$ at the i -th step, and compare it to the calibration value $(BV/TV)_{Calibr}$. If they do not coincide, the new value $Tb.Sp.Factor$ is calculated according to the following formula:

$$(Tb.Sp.Factor)_{i+1} = \frac{(Tb.Sp.Factor)_i}{2} \cdot \left(\frac{(BV/TV)_i}{(BV/TV)_{Calibr}} + 1 \right),$$

where i is the index of the calibration step.

When the BV/TV ratio calculated for the current model falls within the admissible range, the obtained model is considered calibrated, and the associated characteristics of the trabecular structure can be used for further calculations.

5. Model Voxelization and Cortical Layer Description

The final simulation result should be presented in the voxel form for further use in dosimetric calculations and visualization. The subdivision into elementary volumes (voxels) is carried out by a grid of planes located at equal distances from each other and perpendicular to each of the coordinate axes. The distance between the planes corresponds to voxel resolution.

We assume that each voxel contains one of the four homogeneous fillings:

- (1) bone marrow;
- (2) bone substance of trabeculae;
- (3) bone substance of the cortical layer;
- (4) empty space.

Thus, the simulated body is a Boolean four-dimensional array of voxels, represented by three coordinates corresponding to one of the corners of the cube-voxel, and a value from 1 to 4 corresponding to filling the voxel.

Unlike the analytically modeled trabecular structure and surfaces corresponding to the shape of the bone, the cortical layer model is created during the voxelization process. For a voxel partition in the volume of the space on which the stochastic model of the trabeculae (VK) is defined, we construct a parallelepiped (P), in which the modeled figure (F) is fully contained and for which the lengths of its edges are multiples of the voxel resolution $V_F \leq V_P \leq V_K$. Here V_F, V_P, V_K are the volumes of the respective figures. We determine whether a voxel belongs to a particular structure based on the central point of the voxel via the following steps:

1. If the coordinate of the center point of the voxel does not belong to the space bounded by the surfaces of the figure F , then the voxel is assigned material 4.
2. Otherwise, we determine how far the center point is located from the nearest face of F . If the distance is less than or equal to the thickness of the cortical bone, then material 3 is assigned to the voxel.
3. If the previous steps do not yield a result, material 2 is selected if the central point belongs to the space corresponding to the elements of the rod-like structure of the trabeculae; and material 1 if it does not.

Fig. 7 shows the result of the voxelization of the rib model, previously described as an example (see Fig. 3). In this example, no empty space was generated. Bone structures (materials 3 and 2) are shown in black. Bone marrow (material 1) is shown in white.

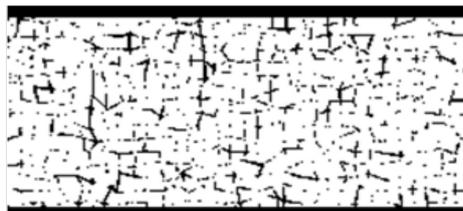


Fig. 7. Visualization of the horizontal section of a rib segment

6. Simulation of Individual Variability

Simulation of individual variability is achieved by perturbing the input parameters of the model⁵. For this purpose, we construct a number of models with randomly generated main parameters based on the population-average macro-dimensions, *Tb.Th*, calibrated *Tb.Sp* and their person-to-person variability (namely, user-defined literature-based standard deviations). The calibration factor (*Tb.Sp.Factor*) for all of these models is the same. Each of the resulting models is subjected to verification; here the calculated *BV / TV* ratio must fall into the admissible range.

The drawing of micro- and macro-parameters is carried out with lognormal and normal approaches respectively, within 90% of the confidence interval. The values must *agree* as follows:

1. If ξ_1, ξ_2 are the random variables describing the *macro* parameters of the model, and ξ_1 takes a value x_1 , then the value $\xi_2 = x_2$ *agrees* with the value $\xi_1 = x_1$ if $F_{\xi_2}(x_2) = F_{\xi_1}(x_1)$. Here $F_{\xi}(x)$ is the cumulative distribution function of the random value ξ . The value can be easily obtained as $x_2 = F_{\xi_2}^{-1}(F_{\xi_1}(x_1))$.

2. If ξ_1, ξ_2 are the random variables describing the *micro* parameters (*Tb.Th* and *Tb.Sp*) of the model, and ξ_1 takes a value x_1 , then value $\xi_2 = x_2$ *agrees* with the value $\xi_1 = x_1$ if $F_{\xi_2}(x_2) = 1 - F_{\xi_1}(x_1)$. Here $F_{\xi}(x)$ is the cumulative distribution function of the random value ξ . The value can be easily obtained as $x_2 = F_{\xi_2}^{-1}(1 - F_{\xi_1}(x_1))$.

The Box – Muller [20] algorithm was used to draw the normal and lognormal random variables.

Conclusion

Originally, parametric stochastic model of bone geometry was developed to be applied for the radiation risk assessments in the South Urals. Radioactive contamination in the South Urals in the 1950s led to radiation exposure to people who were subsequently included in epidemiological cohorts to investigate radiation risk. Among the radionuclides which contaminated the environment, affecting the residents along the Techa riverside and in the East Urals Radioactive Trace, Strontium isotopes were the most dangerous for bone marrow exposure. For example, a statistically significant relationship between the radiation dose and leukemia cases was found in the Techa River cohort [17]. Thus, the assessment of the doses from the incorporated ^{89,90}Sr, aimed at improving the Dosimetric System of the River Techa (TRDS) [18], is of paramount importance in the dosimetric studies currently under way in the JCCRER 1.1 project.

The model presented in this paper will be used to describe the geometry of hematopoietic sites, which in turn will serve as the basis for calculating the doses of irradiation of the hematopoietic cells of the bone marrow.

Currently, work is under way to implement the presented algorithm in the form of a computer programme with an interface designed for radiation biophysicists. The output will be compatible with the existing programme for modelling the transport of emissions.

⁵Note that when constructing a set of models, the macro parameters of the bone (linear dimensions) are subject to the normal distribution. Macro-parameters are assumed to be connected. (If one of the parameters turned out to be higher than the average, then for the others we expect values greater than the average.) Cortical thickness is an independent parameter. When modelling the variants of the spongy structure, its parameters (trabecular thickness and trabecular space) are also related: the greater the thickness of the trabeculae, the less the trabecular separation.

Besides incident-specific study, the model will allow estimating dosimetric uncertainties due to individual variability of bone geometry for the first time. Moreover, the results obtained with the model will be useful for verification of complex CT- and μ CT- based models of ICRP [6–8].

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ПАРАМЕТРИЧЕСКАЯ СТОХАСТИЧЕСКАЯ МОДЕЛЬ ГЕОМЕТРИИ КОСТИ

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Целью настоящего исследования является разработка алгоритма параметрического моделирования костей с учетом особенностей их микроархитектуры, который позволяет генерировать фантомы гематопозитических сегментов кости на основе литературных данных о микро- и макро- размерах сегментов. Мы предлагаем подход,

позволяющий моделировать кости путем их разбиения на небольшие сегменты, описываемые геометрическими фигурами простой формы, заполненные стохастически генерируемой стержнеподобной моделью трабекулярной структуры, с подходящим разрешением воксела. Предлагаемый метод позволяет избежать недостатков непараметрического моделирования, основанного на индивидуальной компьютерной томографии. Параметрический подход позволяет, также, моделировать индивидуальную изменчивость костно-специфических размеров. Модель, представленная в этой статье, может быть использована для описания геометрии гемопоэтических сайтов, которые, в свою очередь, служат основой для расчета доз облучения гемопоэтических клеток костного мозга от инкорпорированных бета-излучателей.

Ключевые слова: микро- и макро- структура трабекулярной кости; стохастическое моделирование; вокселлизация.

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